

## GENERAL DESCRIPTION

Porcine Circovirus type 2(PCV2) has been identified as the causal agent of post weaning multisystemic wasting syndrom (PMWS). PCV is a single-stranded DNA virus (class II), that is nonenveloped with an unsegmented circular genome.

HRPro® PCV2 AB ELISA is quantitative antibody ELISA test kit for PCV type 2. HRPro® PCV2 AB ELISA is an indirect ELISA and this quantitative serological test may supply the useful information of immunoreactivity level of herds by checking average antibody titer of herds. HRPro® PCV2 AB ELISA was developed to detect anti-PCV2 antibodies in serum using specific antigen of PCV2.

HRPro® PCV2 AB ELISA is excellent reagent to herd based antibody test instead of individual test by analysis of positive rates, average titer, seroconversion.

## KIT COMPONENTS

| Reagents                         | 480 tests |
|----------------------------------|-----------|
| ① PCV2 Ag coated plate           | 5 plates  |
| ② 10X Washing Buffer             | 240ml X 1 |
| ③ Dilution Buffer                | 240ml X 1 |
| ④ HRPO Anti -Swine IgG Conjugate | 70ml X 1  |
| ⑤ Positive Control, PC           | 7.0ml X 1 |
| ⑥ Negative Control, NC           | 7.0ml X 1 |
| ⑦ TMB Substrate                  | 70ml X 1  |
| ⑧ Stop Solution                  | 40ml X 1  |
| ⑨ Sealing Film                   | 5 sheets  |
| ⑩ Instruction Manual             | 1 copy    |

## PREPARATION

- All reagents must be allowed to come to room temperature (20~25 °C) before use. Mix reagents by gentle swirling.
- 1X washing buffer preparation
  - Shake 10X Washing Buffer(②) gently.
  - Dilute 1 part of 10X Washing Buffer(②) with 9 parts of deionized water. The diluted 1X washing buffer is stable for 2 weeks at room temperature (20~25 °C).
- Sample preparation
  - Use the serum samples as fresh as possible
  - Use fresh samples for the best result. Serum samples can be stored at 2~8°C for less than 3 days or -20 °C for a longer period. Do not freeze and thaw serum samples repeatedly. **Sera with hemolysis or bacterial contamination are not suitable for the analysis!**
  - Visible solid materials in serum samples should be separated by centrifugation.

- Sample dilution
  - Prepare the 1 ml deep-well-plate (DWP, 96-well, not offered) or suitable tubes.
  - Dilute the sample 1/100 in dilution buffer(③). Add 495 µl dilution buffer(③) to each well of DWP and add 5 µl serum to each well. Diluted Samples should be mixed prior to dispensing into the Antigen Coated Plate(①).
- Do not dilute the Positive Control (PC, ⑤) and the Negative Control (NC, ⑥).**
- TMB Substrate (⑦) should be warmed up for 30 minutes at room temperature (20~25 °C) before use (10ml/plate). If stored at low temperature, the color development may be poor.

## TEST PROCEDURE

- Remove the Antigen Coated Plate (①) from protective foil pouch.**
- Dispense 100µl of diluted sample into appropriate wells. **Use care not to spill samples from well to well.**
- Dispense 100µl of undiluted NC( ⑥ ) and undiluted PC(⑤) into duplicate wells.
- Seal the plate and Incubate for 30 minutes at 25 ±1.0 °C.
- Wash each well 3 times with 1X washing buffer (300µl per well). And remove contents of well at each washing steps. After final washing, hit plate to the paper towel briefly to remove solution.
- Dispense 100µl of HRPO Anti-Swine IgG Conjugate(④) to each well.
- Seal the plate and incubate for 30 minutes at 25±1.0 °C.
- Wash each well as step 5.
- Dispense 100µl of TMB substrate (⑦) to each well.
- Seal the plate and incubate for 15 minutes at 25 ±1 °C. Check the density of color development by naked eyes.
- Add 50µl of Stop Solution (⑧) to each well of the plate. Shake the test plate shortly (5~10 sec). **Be careful not to spill.**
- Measure and record the A (450nm) for samples and controls immediately.
- Validate and calculate the results.

Plate template example (1-well Test )

|   | 1 | 2  | 3  | 4  | 5  | 6  | 7  | 8  | 9  | 10 | 11 | 12 |
|---|---|----|----|----|----|----|----|----|----|----|----|----|
| A | 1 | 9  | 17 | 25 | 33 | 41 | 49 | 57 | 65 | 73 | 81 | 87 |
| B | 2 | 10 | 18 | 26 | 34 | 42 | 50 | 58 | 66 | 74 | 82 | 88 |
| C | 3 | 11 | 19 | 27 | 35 | 43 | 51 | 59 | 67 | 75 | 83 | 89 |
| D | 4 | 12 | 20 | 28 | 36 | 44 | 52 | 60 | 68 | 76 | 84 | 90 |
| E | 5 | 13 | 21 | 29 | 37 | 45 | 53 | 61 | 69 | 77 | 85 | 91 |
| F | 6 | 14 | 22 | 30 | 38 | 46 | 54 | 62 | 70 | 78 | 86 | 92 |
| G | 7 | 15 | 23 | 31 | 39 | 47 | 55 | 63 | 71 | 79 | PC | PC |
| H | 8 | 16 | 24 | 32 | 40 | 48 | 56 | 64 | 72 | 80 | NC | NC |

## RESULT INTERPRETATION

1. Validate if the mean OD of the PC is higher than 0.50 and the mean OD of the NC is lower than 0.30. If these criteria are not met, the test are invalid and the samples must be retested.
2. Calculate the sample to positive ratio (S/P ratio) by following the formula

$$\text{S/P ratio} = \frac{\text{Sample OD} - \text{NC mean OD}}{\text{PC mean OD} - \text{NC mean OD}}$$

3. Result interpretation
  - 1) Test samples having  $\geq 0.4$  S/P ratio are positive.
  - 2) Test samples having  $0.3 \leq \text{S/P ratio} < 0.4$  are Suspects.
  - 3) Test samples having  $< 0.3$  S/P ratio negative.

| S/P value                         | Interpretation |
|-----------------------------------|----------------|
| $\text{S/P} \geq 0.4$             | Positive       |
| $0.3 \leq \text{S/P ratio} < 0.4$ | Suspect        |
| $\text{S/P} < 0.3$                | Negative       |

❖PCV2- AB can't differentiate the antibody level of infection and vaccination.

4. Example of result calculation and interpretation
  - 1) ODs of NC : 0.085, 0.091  
Mean OD =  $(0.085 + 0.091) / 2 = 0.088$  (valid)
  - 2) ODs of PC :1.121, 1.201  
Mean OD =  $(1.121 + 1.201) / 2 = 1.161$  ( valid)
  - 3) OD of Sample : 0.431  
S/P ratio of the sample =  $(0.431 - 0.088) / (1.161 - 0.088) = 0.319$
  - 4) Result interpretation: Suspect

## PRECAUTIONS

1. All reagents must be allowed to come to room temperature (20~25 °C) before use. Mix reagents by gentle swirling. After use, return to 2~8 °C.
2. Read this instruction manual thoroughly and follow all steps strictly for successful use of the product.
3. All test samples should be considered potentially infectious and all items contacting the samples should be considered contaminated.
4. Do not use expired or contaminated reagents.
5. Do not use reagents from other kits or lots.
6. Do not mix reagents from different lots of this same product.
7. Do not expose the reagents to excessive heat or direct light during storage and incubation.
8. Incomplete washing adversely may affect the result and precision of the assay.
9. Avoid microbial contamination of the reagents.
10. Avoid contamination of the TMB Substrate(⑦) with the HRPO Anti-Swine IgG Conjugate(④).
11. Wear personal protective equipment (PPE) such as lab coat, goggle, and disposable gloves while performing the assay. Wash hands thoroughly afterwards.
12. Do not eat, drink, smoke or apply cosmetics where kit reagents are handled. Do not pipette by mouth.
13. Pipette tips must be changed after each pipetting step. Use a clean disposable pipette tip for all steps.
14. Use care not to spill samples from well to well.
15. Deionized water or equal must be used to prepare the washing buffer.
16. Unused strips should be stored in the sealed foil pouch at 2~8 °C. Re-sealed strips are recommended to use within one week.
17. For veterinary use only.

## STORAGE AND STABILITY

Store all reagents at 2~8 °C. Do not freeze. Reagents remain stable until the expiration date when stored as instructed.

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## QUICK PROTOCOL

Serum Dilution (Dilution factor 1:100)  
Dilution buffer 495  $\mu\text{l}$  + Serum 5  $\mu\text{l}$

PCV2 Ag Coated Plate

Add 100  $\mu\text{l}$  of diluted Samples  
&  
Add 100  $\mu\text{l}$  of Undiluted PC, NC



25°C, 30 min

Washing  
3 times

Dispense 100  $\mu\text{l}$  of  
HRPO Anti-Swine IgG Conjugate



25°C, 30 min

Washing  
3 times

Dispense 100  $\mu\text{l}$  of  
TMB Substrate



25°C, 15 min

Dispense 50  $\mu\text{l}$  of  
Stop Solution

Measure OD at 450nm